

# **DOE Bioenergy Technologies Office (BETO) 2021 Project Peer Review**

## **ABF Industry Engagement Lab Call - Visolis: Production of High-Value Chemicals from Renewable Feedstocks**

**3/10/2021**

**ABF**

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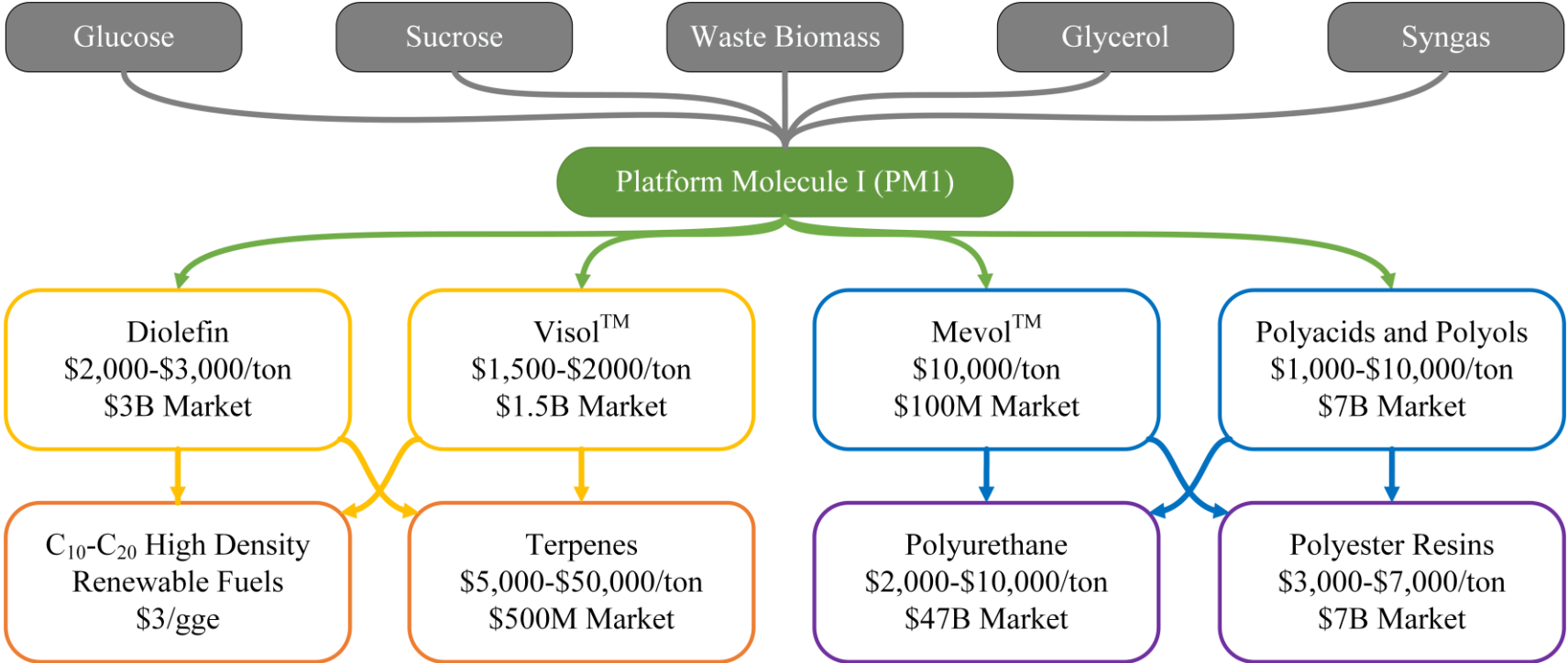
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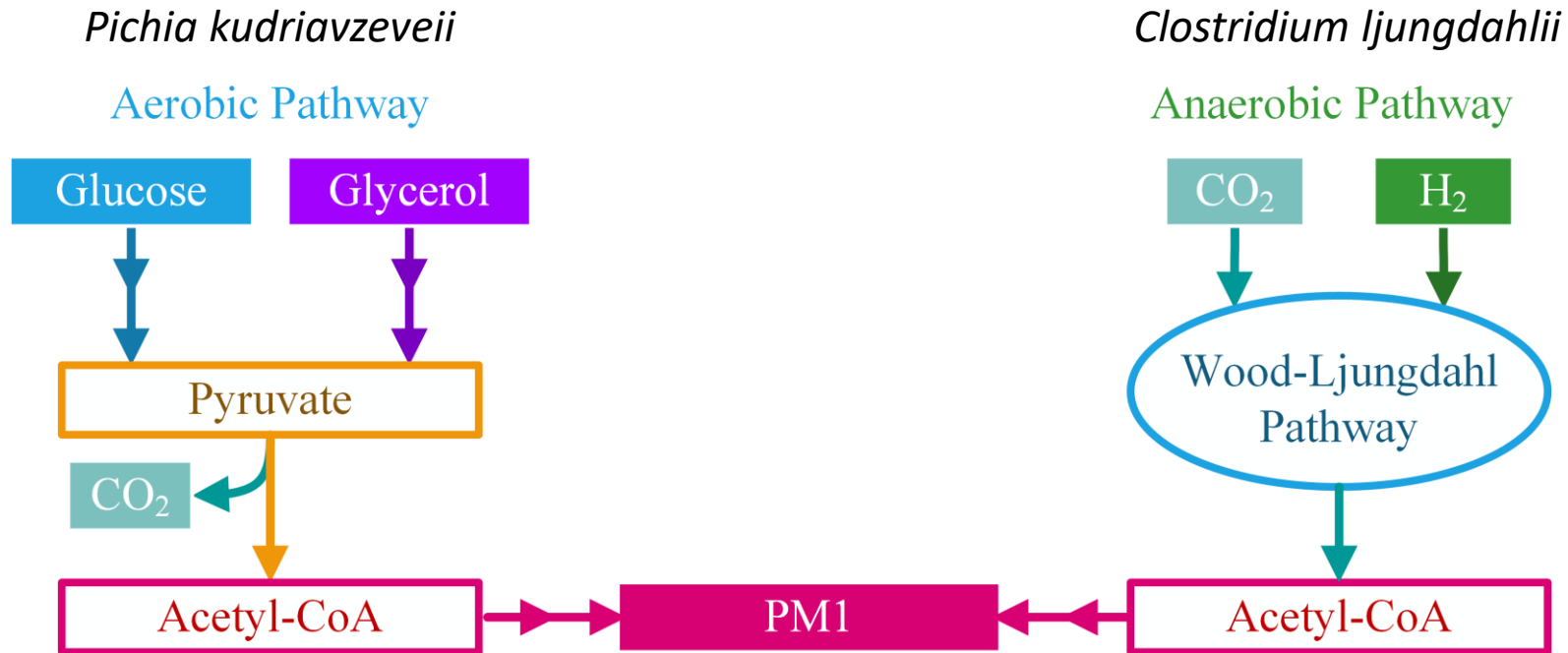
# Presentation Outline

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# Platform molecule production from diverse feedstocks



# Non-conventional hosts with potential for performance advantage



**A New Paradigm:** a hybrid biomass-conversion technology integrating thermochemical gasification with syngas fermentation (CO/H<sub>2</sub>/CO<sub>2</sub>) to improve biorefinery economics

- Lower capital and operating cost with low pH fermentation
- Better biomass utilization
- Higher carbon efficiency (compared to carbon loss from glycolysis)

# Project Structure

Improving genetic tools  
for *C. ljungdahlii*



Metabolic Engineering of  
*C. ljungdahlii*



Metabolic Engineering  
of *P. kudriavzevii*



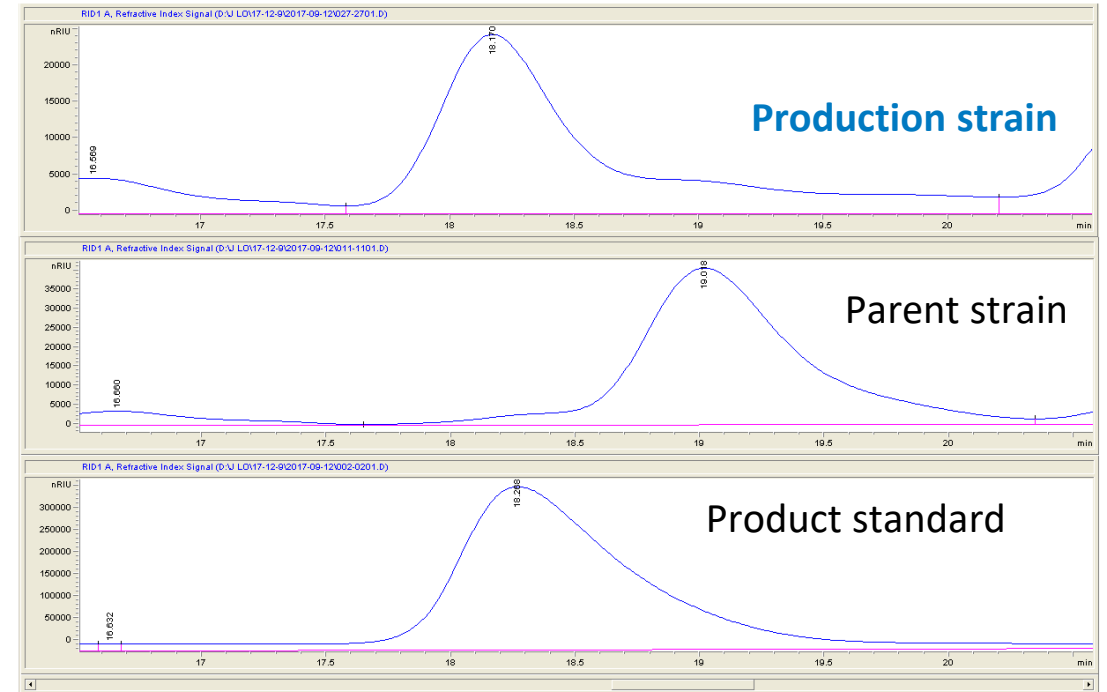
Bioreactor  
Experiments/TEA



# Engineering PM1 production in *C. ljungdahlii*

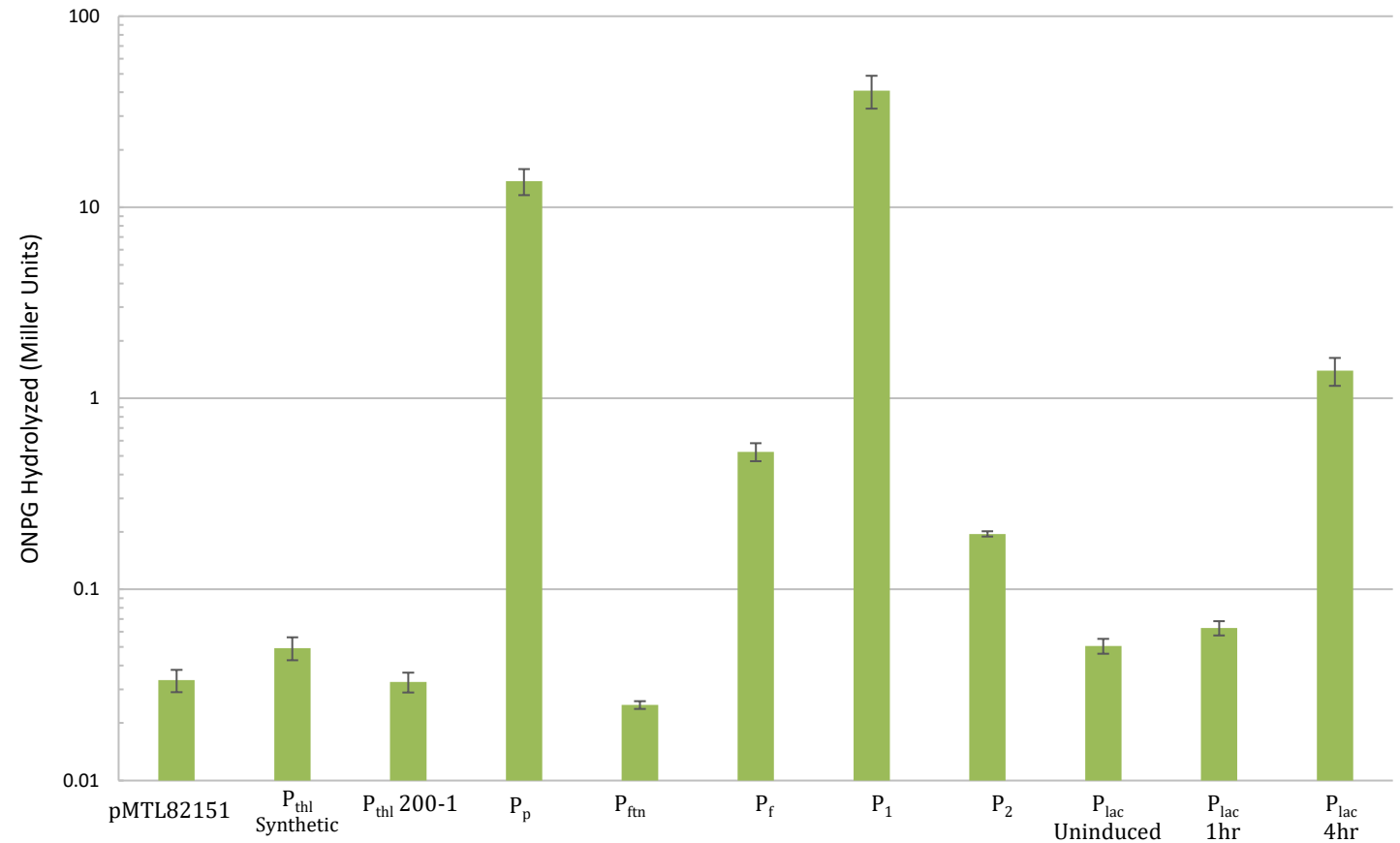
- **Higher Carbon Efficiency:** CO<sub>2</sub>-fixing autotrophic microbes such as *Clostridium ljungdahlii* metabolize CO<sub>2</sub>, H<sub>2</sub>, and CO yielding acetyl-CoA.
- Acetyl-CoA upgrade leads to PM1, a precursor with myriads of applications.
- Initially transformed production related constructs m<sub>1</sub> and m<sub>2</sub> into *C. ljungdahlii* using a replicating plasmid:
- Used *thl* and *araE* promoters to drive expression of genes
- Detected ~80 mg/L PM1 in WT background strain

## *C. ljungdahlii*



# Engineering PM1 production in *C. ljungdahliae*

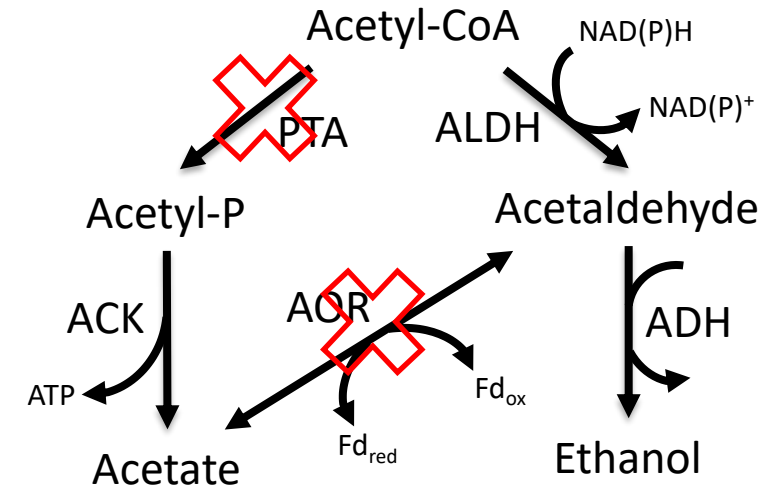
- Characterized promoters that span ~1000-fold range of expressions
- Confirmed trends with new anaerobic fluorescent marker (Y-FAST)
- Identified a promoter that is stronger than ones currently used



# Engineering PM1 production in *C. ljungdahlii*

Using newly characterized promoters, we generated new construct with stronger promoters to drive production genes:

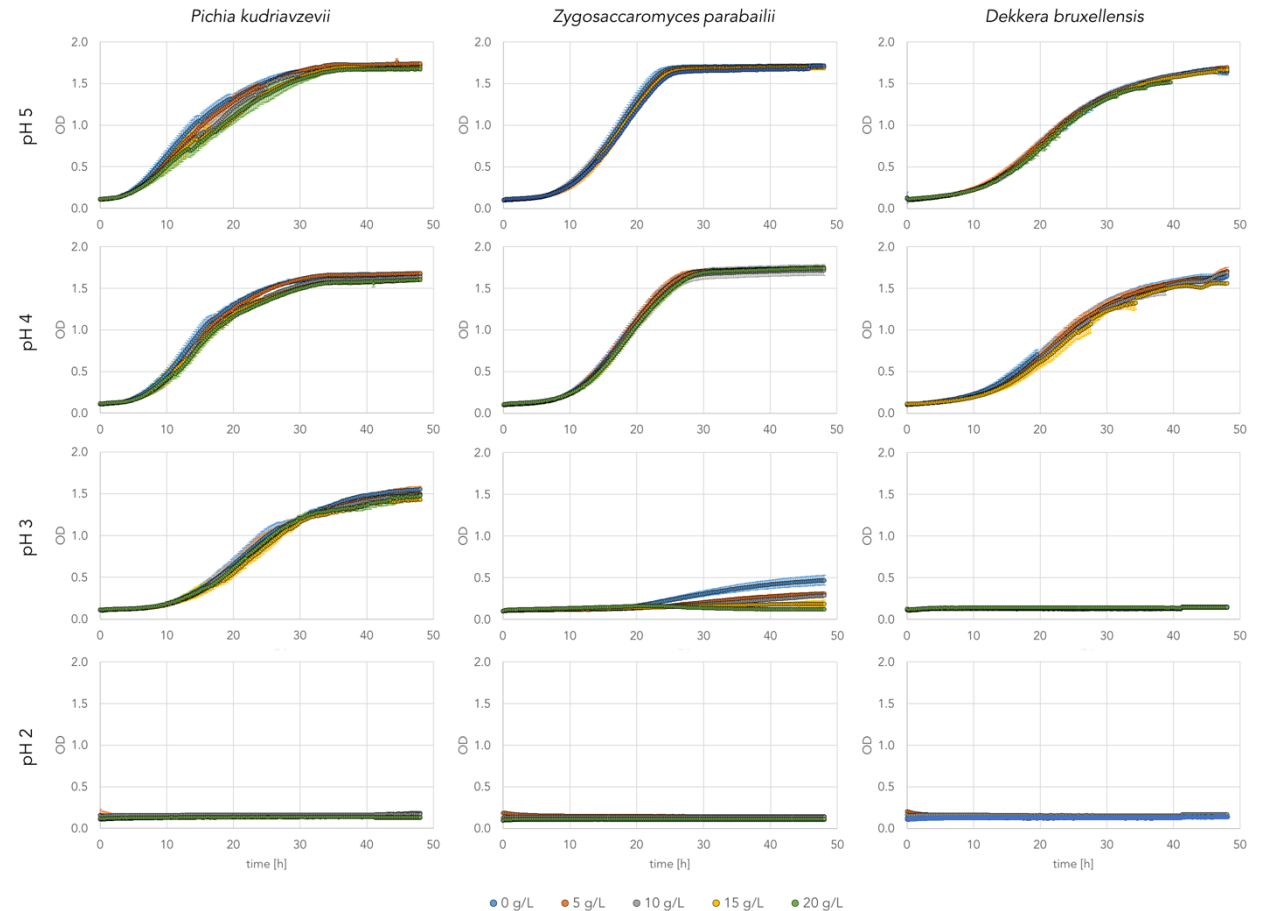
- We put this construct into a  $\Delta pta \Delta aor2$  strain that has reduced acetate
- Increased PM1 production to  $\sim 700$  mg/L, up from 80 mg/L
- Provides a proof-of-concept that PM1 production via a synthetic pathway is linked to native cellular metabolic network in chassis strain.



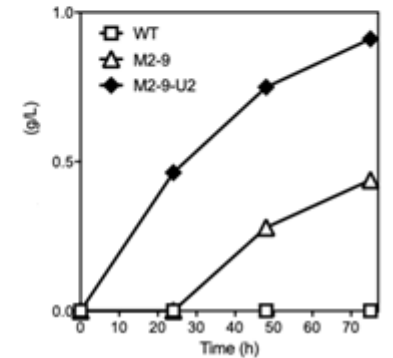
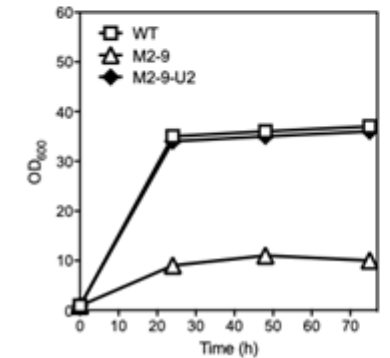
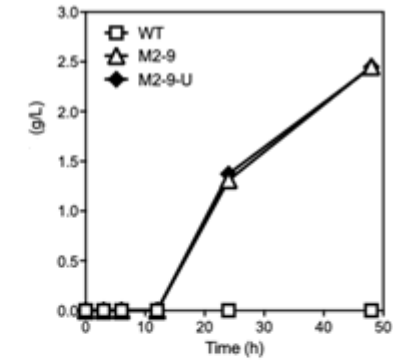
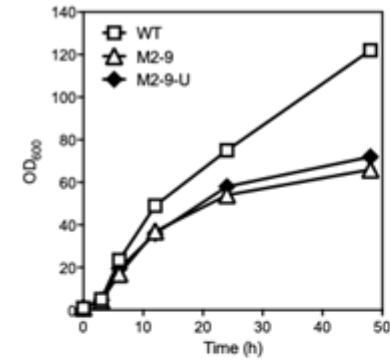
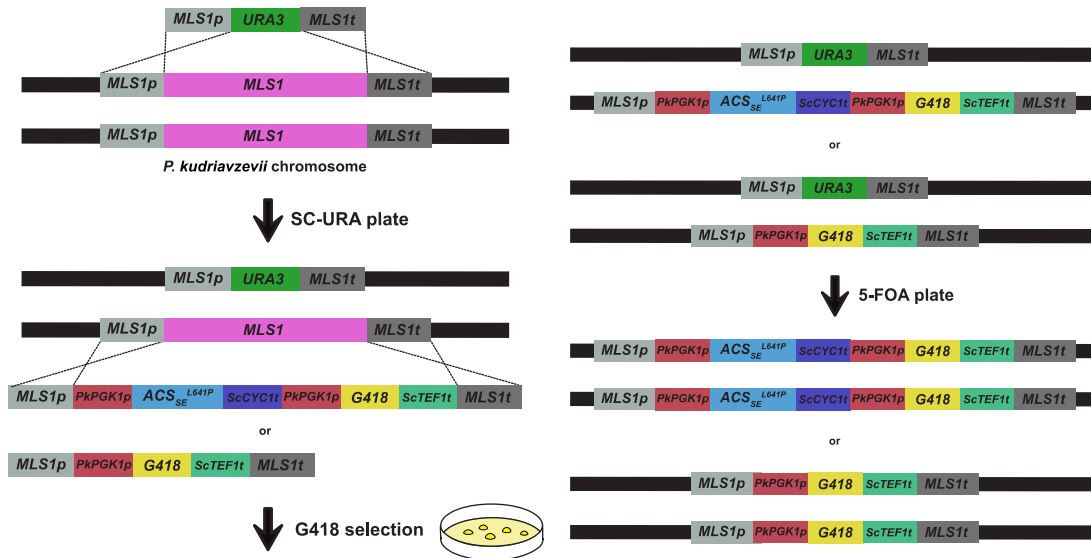


# Engineering PM1 production in a low pH yeast

- Cultivations of *P. kudriavzevii*, *Z. parabailii*, and *D. bruxellensis* were performed in the presence of 0, 5, 10, 15 g/L PM1 at initial pH of 5.0, 4.0, 3.0 and 2.0.
- While *Z. parabailii* and *D. bruxellensis* exhibited no growth at pH 3, *P. kudriavzevii* growth profiles were largely unaffected, confirming that this yeast is a suitable chassis for PM1 production under low pH conditions.

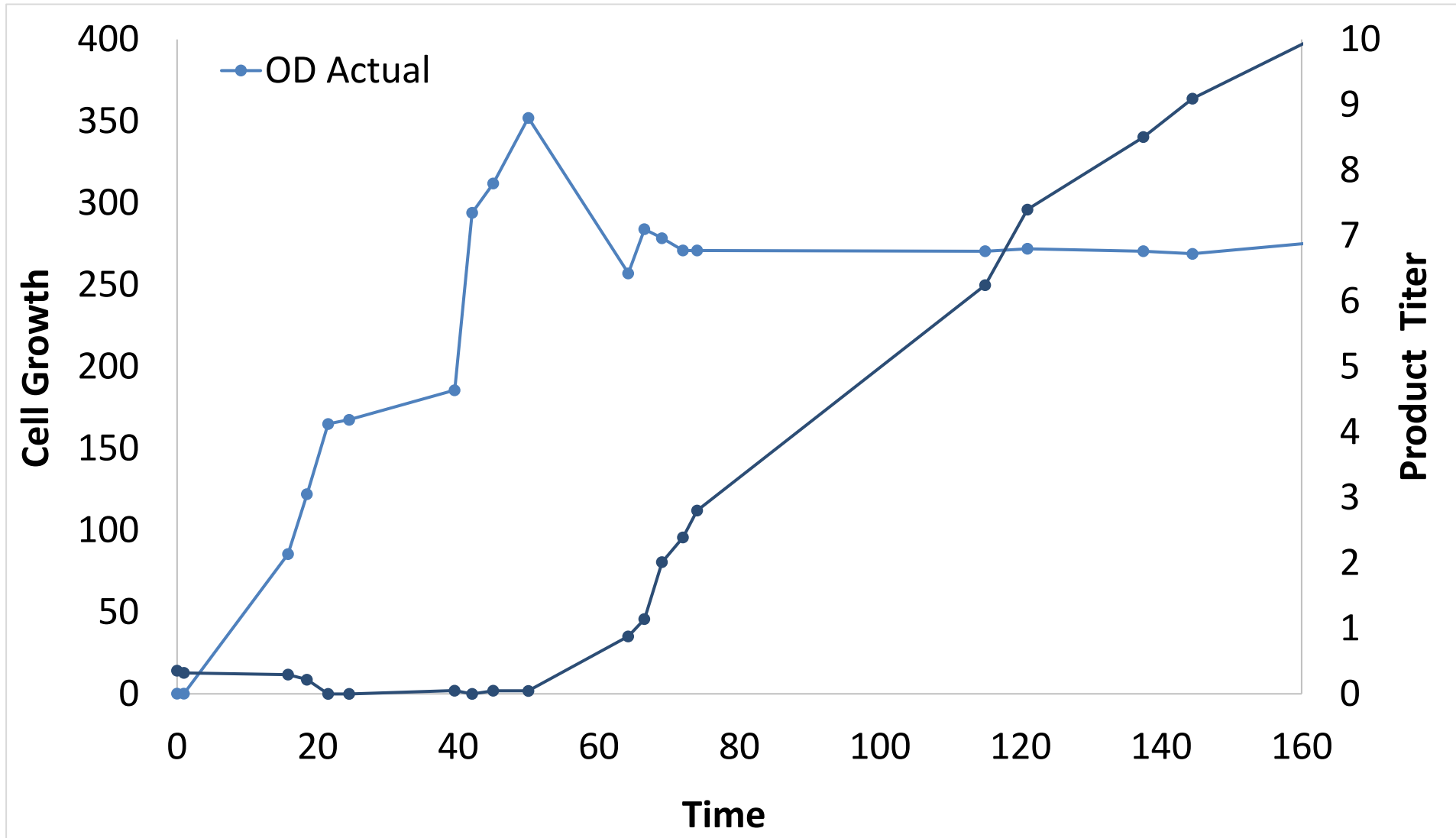


# Engineering PM1 production in a low pH yeast



- Using URA3 as a selection/counterselection system, the following genetic edits were introduced into the low pH yeast *Pichia kudriavzevii*:
- Extra copies of PM1 production genes on both chromosomes to increase production
- Deletion of competing malate pathway gene MLS1 by replacement with URA3 to improve growth at low pH

# Engineering PM1 production in a low pH yeast



# Progress and Outcomes- Summary and Impact

## *C. Ljungdahlii* chassis

- Improved transformation protocol for *C. ljungdahlii*.
- Initial *C. ljungdahlii* strain with PM1 production validated.
- Construction of promoter libraries for *C. ljungdahlii*.
- *Development of genomic integration protocol for C. ljungdahlii.*

## *P. kudriavzevii* chassis

- Improved pH tolerance of *P. kudriavzevii* for PM1 production.
- DBTL cycle to produce a *P. kudriavzevii* strain with PM1 production.
- Demonstration of PM1 production at 3X project target in bench top bioreactors at low pH

# Quad Chart Overview

## Timeline

- March 1, 2018
- Sept 30, 2020

	FY20	Project
DOE Funding	(10/01/2019 – 9/30/2020)	\$0  \$500,000 (project)

## Project Partners

- National Renewable Energy Lab
- Oakridge National Lab

## Barriers addressed

1. Develop cost-effective biological synthesis technologies.

## Project Goal

Demonstrate use of non-traditional hosts for production of platform molecule

## End of Project Milestone

Developed tools for engineering or two non-traditional hosts and demonstrated production of key platform molecule

## Funding Mechanism

ABF Industry Engagement Lab Call